



## EFFECT OF PUMPKIN (*CUCURBITA PEPO* L.) AND MARIGOLD (*TAGETES PATULA* L.) EXTRACTS ON HIPPOCAMPAL MITOCHONDRIA FUNCTIONAL ACTIVITY WITHIN CONDITIONS OF EXPERIMENTAL ACUTE BRAIN HYPOMETABOLISM

A.V. Voronkov, D.I. Pozdnyakov, S.L. Adzhiakhmetova, N.M. Chervonnaya,  
K.A. Miroshnichenko, A.V. Sosnovskaya, E.I. Chereshkova

Pyatigorsk Medical and Pharmaceutical Institute – branch of Volgograd State Medical University  
11, Kalinin Ave., Pyatigorsk, Russia, 357532

E-mail: pozdniackow.dmitry@yandex.ru

Received 9 July 2019

Review (1) 31 July 2019

Review (2) 8 August 2019

Accepted: 15 August 2019

**The aim** of the study is to evaluate the effect of pumpkin (*Cucurbita pepo* L.) and marigold extracts (*Tagetes patula* L.) on the hippocampal mitochondria functional activity within the conditions of experimental acute brain hypometabolism.

**Materials and methods.** The work was performed on 50 male Wistar rats, which reproduced an acute brain hypometabolic state by administration of a 3M sodium azide solution in hippocampus ( $n = 40$  and  $n = 10$  – a group of sham-operated animals). The test extracts and the reference drug – EGb 761 – were prophylactically administered at the dose of 100 mg/kg *per os* for 10 days. 24 hours after the last administration, sodium azide was injected, the brain was taken, the hippocampus was isolated to obtain a supernatant and determine the parameters of mitochondrial respiration, the intensity of anaerobic processes, the concentration of the apoptosis-inducing factor, endonuclease G, and  $\beta$ -amyloid.

**Results.** The carried out study established that the prophylactic administration of pumpkin and marigold extracts contributed to the restoration of a mitochondrial function and a decrease in the intensity of anaerobic processes. In the group of the rats treated with pumpkin and marigold extracts, an increase of ATP concentration in the hippocampal supernatant by 65.7% ( $p < 0.002$ ) was observed; it was 66.2% ( $p < 0.002$ ) relative to the animals deprived of pharmacological support. When the rats were treated with pumpkin and marigold extracts, a decrease in the concentration of apoptosis-inducing factor (by 33% ( $p < 0.002$ ) and 38.3% ( $p < 0.002$ ), respectively) and endonuclease G (by 3.6 times ( $p < 0.002$ ) and 4.4 times ( $p < 0.002$ ), respectively) was also noted. The administration of pumpkin and marigold extracts reduced the amyloid  $\beta$ -peptide concentration in the rats' hippocampus by 54.4% ( $p < 0.0002$ ) and 54.4% ( $p < 0.0002$ ), respectively. The test-extracts had an equivalent therapeutic efficacy with the reference drug.

**Conclusion** On the basis of the obtained data, it is possible to suggest the prospect of a further study of pumpkin and marigold extracts as the drugs of a targeted correction of cerebral hypometabolism.

**Keywords:** plant extracts, hypometabolism, hippocampus, mitochondria

**For citation:** A.V. Voronkov, D.I. Pozdnyakov, S.L. Adzhiakhmetova, N.M. Chervonnaya, K.A. Miroshnichenko, A.V. Sosnovskaya, E.I. Chereshkova. Effect of pumpkin (*cucurbita pepo* L.) and marigold (*tagetes patula* L.) extracts on hippocampal mitochondria functional activity within conditions of experimental acute brain hypometabolism. *Pharmacy & Pharmacology*. 2019;7(4): 198-207. DOI: 10.19163/2307-9266-2019-7-4-198-207

©А.В. Воронков, Д.И. Поздняков, С.Л. Аджиахметова, Н.М. Червонная, К.А. Мирошниченко, А.В. Сосновская, Е.И. Шерешкова, 2019

**Для цитирования:** А.В. Воронков, Д.И. Поздняков, С.Л. Аджиахметова, Н.М. Червонная, К.А. Мирошниченко, А.В. Сосновская, Е.И. Шерешкова. Влияние экстракта тыквы обыкновенной (*cucurbita pepo* L.) и экстракта бархатцев распростертых (*tagetes patula* L.) на функциональную активность митохондрий гиппокампа в условиях экспериментального острого гипометаболизма головного мозга. *Фармация и фармакология*. 2019;7(4): 198-207. DOI: 10.19163/2307-9266-2019-7-4-198-207

## **ВЛИЯНИЕ ЭКСТРАКТА ТЫКВЫ ОБЫКНОВЕННОЙ (CUCURBITA PEPO L.) И ЭКСТРАКТА БАРХАТЦЕВ РАСПРОСТЕРТЫХ (TAGETES PATULA L.) НА ФУНКЦИОНАЛЬНУЮ АКТИВНОСТЬ МИТОХОНДРИЙ ГИППОКАМПА В УСЛОВИЯХ ЭКСПЕРИМЕНТАЛЬНОГО ОСТРОГО ГИПОМЕТАБОЛИЗМА ГОЛОВНОГО МОЗГА**

**А.В. Воронков, Д.И. Поздняков, С.Л. Аджирахметова, Н.М. Червонная,  
К.А. Мирошниченко, А.В. Сосновская, Е.И. Шерешкова**

Пятигорский медико-фармацевтический институт – филиал ФГБОУ ВО «Волгоградский  
государственный медицинский университет» Минздрава России  
Россия, 357532, г. Пятигорск, пр. Калинина, д. 11

E-mail: pozdniackow.dmitry@yandex.ru

Получено 09.07.2019

Рецензия (1) 31.07.2019

Рецензия (2) 08.08.2019

Принята к печати 15.08.2019

**Цель исследования.** Оценить влияние экстракта тыквы обыкновенной (*Cucurbita pepo* L.) и экстракта бархатцев распростертых (*Tagetes patula* L.) на функциональную активность митохондрий гиппокампа в условиях экспериментального острого гипометаболизма головного мозга.

**Материалы и методы.** Работа выполнена на 50 крысах-самцах линии *Wistar*, которым моделировали острое гипометаболическое состояние путем введения 3М раствора натрия азида в гиппокамп ( $n=40$  и  $n=10$  – группа ложнопериоперированных животных). Исследуемые экстракты и препарат сравнения – EGb 761 вводили в дозе 100 мг/кг *per os*, профилактически на протяжении 10 дней. Через 24 часа после введения натрия азида осуществляли забор головного мозга, выделяли гиппокамп с получением супернатанта и определением в нем параметров митохондриального дыхания, интенсивности анаэробных процессов, концентрации апоптоз-индуцирующего фактора, эндонуклеазы G и  $\beta$ -амилоида.

**Результаты исследования.** В ходе исследования было установлено, что профилактическое введение изучаемых экстрактов тыквы и бархатцев способствовало восстановлению митохондриальной функции и снижению интенсивности анаэробных процессов. При этом в группах крыс, получавших экстракт тыквы и бархатцев, отмечено повышение концентрации АТФ в супернатанте гиппокампа относительно животных без фармакологической поддержки на 65,7% ( $p<0,002$ ) и 66,2% ( $p<0,002$ ); соответственно. Также при введении крысам экстрактов тыквы и бархатцев наблюдалось снижение концентрации апоптоз-индуцирующего фактора (на 33 % ( $p<0,002$ ) и 38,3% ( $p<0,002$ ) соответственно) и эндонуклеазы G (в 3,6 раза ( $p<0,002$ ) и в 4,4 раза ( $p<0,002$ ) соответственно). При применении исследуемых экстрактов тыквы и бархатцев отмечено снижение концентрации амилоидного  $\beta$ -пептида в гиппокампе крыс на 54,4% ( $p<0,0002$ ) и 54,4% ( $p<0,0002$ ) соответственно. При этом исследуемые экстракты обладали эквивалентной терапевтической эффективностью с препаратом сравнения.

**Заключение.** На основании полученных данных можно предположить перспективность дальнейшего изучения экстрактов тыквы и бархатцев как средств целенаправленной коррекции церебрального гипометаболизма.

**Ключевые слова:** растительные экстракты, гипометаболизм, гиппокамп, митохондрии

### **INTRODUCTION**

Alzheimer's disease (AD) is a worsening neurodegenerative disease which accounts for 50–70% of dementia cases, comprising more than 12 million people [1]. Generally, clinical manifestations of AD are observed in the late stages of the disease and are associated with deposition of cytotoxic  $\beta$ -amyloid in the brain structures [2]. Amyloid  $\beta$ -peptide is formed as a result of proteolysis of transmembrane protein – amyloid precursor protein (APP) during catalysis of secretase enzymes. To date, it has been established that the accumulation of  $\beta$ -amyloid is one of the most reliable and early markers of irreversible neurodegeneration [3]. In the processes of neuronal degradation, various  $\beta$ -amyloid isoforms

form two types of cytotoxic conglomerates: the aggregates not stabilized by metal ions (usually soluble in water and having little cytotoxic potential) and associates in which divalent metal ions are present (these conglomerates form covalently cross-linked oligomers, which are then deposited in the cytoplasm of neurons (mainly the hippocampus) in the form of amyloid plaques) [4]. The excess of amyloid  $\beta$ -peptide in brain cells leads to irreversible phosphorylation of tau protein, resulting in the increased production of 4-hydroxynonenal – membrane-toxic aldehyde, which, in its turn, initiates oxidative processes in neuronal membranes, accompanied by a deterioration of the ion transport ATPases, functioning as glucose and glutamate transporters [5]. As a result,

the described processes lead to the impairment in synaptic transmission and AD worsening.

The research group of *Del Prete D, et al., 2017* found out that the amyloid  $\beta$ -peptide precursor protein can be localized on mitochondrial membranes. In addition, a secretase complex, mainly represented by  $\gamma$ -secretase, is found on mitochondrial membranes, which, in turn, provides a rapid synthesis of  $\beta$ -amyloid from its precursor [6].

In a number works it is noted that mitochondrial  $\beta$ -amyloid can play a leading role in the pathological cascade, adducting to early neurons death within AD conditions. Thus, a study by *Lustbader J W, et al., 2004* showed that there is a direct relationship between a mitochondrial dysfunction and a hypometabolic brain state, on the one hand, and an increase of the  $\beta$ -amyloid cytotoxicity, on the other [7]. Further on, *Chen X. et al., 2006* found out that within AD conditions, brain hypometabolism arising with an increase of the amyloid  $\beta$ -peptide concentration is associated with a malfunction of electron transport reactions in the mitochondrial respiratory chain, activation of anaerobic oxidative processes, and intensification of the internal apoptosis pathway (the effector system – endonuclease G) [8]. However, it should be noted that activation of the mitochondrial  $\gamma$ -secretase complex is observed only with a decrease in the intensity of metabolic reactions – a hypometabolic state [9]. Thus, it can be assumed that a targeted pharmacological correction of cerebral hypometabolism may be a promising method for the early treatment of AD. In this regard, the **aim of this study** was to assess the effect of pumpkin (*Cucurbita pepo* L.) and marigold extracts (*Tagetes patula* L.) on the hippocampal mitochondria functional activity within experimental acute brain hypometabolism conditions.

## MATERIALS AND METHODS

### Experimental animals

In the experiment, 50 sexually mature male Wistar rats with a body weight of 220–240 grams) were used. The animals had been obtained from “Rappolovo”, the Federal State Unitary Enterprise “Nursery of laboratory animals (Leningrad Region). During the experiment, the rats were kept under standard vivarium conditions at the air temperature of  $22 \pm 2^\circ\text{C}$ , the relative humidity of  $60 \pm 5\%$  and a natural circadian cycle change. The animals were kept in macrolon cells by 10 individuals, where a granular beech fraction was used as a nesting material. The rats received extruded food and water *ad libitum*. Manipulations with the experimental animals were performed in accordance with the generally accepted ethical standards adopted by the European Convention

for the protection of vertebrate animals used for experimental and other scientific purposes (1986) and taking into account the International recommendations of the European Convention for the protection of vertebrate animals used in experimental studies (1997) [24, 25].

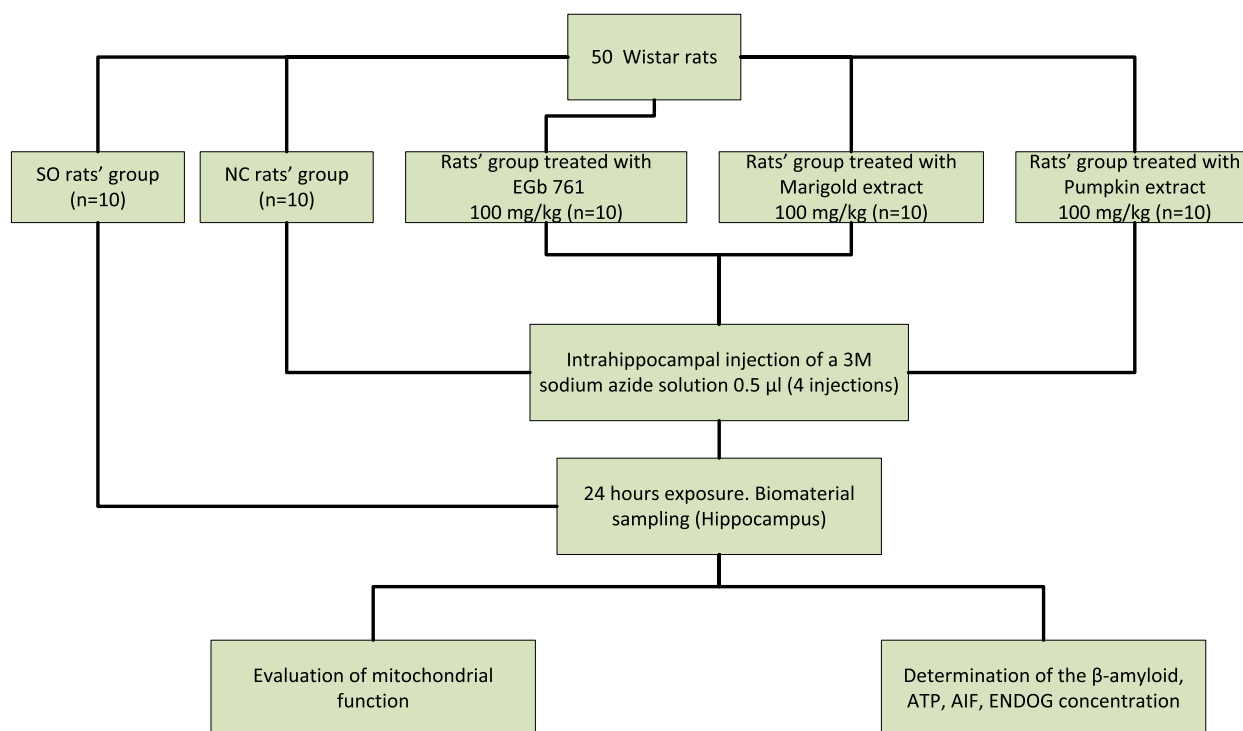
### Test objects. Experiment design

In this work, a thick pumpkin extract (*Cucurbita pepo* L.) and a thick marigold extract (*Tagetes patula* L.) were used as test objects. Marigold inflorescences (*Tagetes patula* L.) of the «Carmen» variety were collected in the botanical garden of Pyatigorsk Medical and Pharmaceutical Institute – branch of Volgograd State Medical University. The pumpkin fruits (*Cucurbita pepo* L.) of the «Atlant srednepozdnyj» variety were collected in the «Aleksandrovskij» district, Stavropol region. The raw materials samples were harvested in the period from June to September 2016–2018. Fresh pumpkin fruits and marigold inflorescences, dried in shadow, were used as raw materials.

EGb 761 (a standardized *Ginkgo biloba* extract manufactured by Hunan Warrant Pharmaceuticals, China) was a reference drug. The reference drug was administered *per os* at the dose of 100 mg/kg [10] before a surgery operation for 10 days. The test objects were administered similarly to the reference drug (prophylactically, at the dose of 100 mg/kg for 10 days). During the experiment, the following experimental groups of animals were formed: sham-operated rats (SO,  $n = 10$ ); a negative control group of rats, deprived of pharmacological support (NC,  $n = 10$ ); groups of animals treated with the reference drug and test-objects ( $n = 10$  in each experimental group). The study design is shown in Fig. 1.

### Experimental model of cerebral hypometabolism

Cerebral hypometabolism was modeled by an intra-hippocampal injection of a 3M sodium azide solution, for which the animals had been anesthetized (chloral hydrate 350 mg/kg, intraperitoneally), the rats' heads were fixed, the pelage was removed and the skull was scalped. Then, in the left and right hemispheres, trepanation holes were drilled with a 1 mm diameter trepanation bur with the stereotactic coordinates P:  $-3.0$ ; ML:  $\pm 3.0$ , V:  $-3.0$  (from Bregma), which corresponded to the dorsal part of the hippocampus [11]. Then, a 30G needle was slowly inserted into the trepanation hole to the depth of 1.5 mm; a 3 M sodium azide solution (pH = 7.4) was injected in 0.5  $\mu\text{l}$  volume, alternating with the right and left hemispheres (totally, there were 4 injections made). The needle was removed after 30 seconds since the insertion. The wound was sutured, the seam treated with a 5% iodine solution. Biomaterial was taken 24 hours after the surgery [12].



**Figure 1 – Experiment design**

Note: SO – a sham-operated rats group; NC – a negative control animals group; ATP – adenosine triphosphate; AIF – apoptosis-inducing factor; ENDOG – endonuclease G.

### Biomaterial sampling

In the work, the rat hippocampus was used as biomaterial. The hippocampus was removed according to the standard procedure. The isolated hippocampus was divided into 2 parts: the first was homogenized in the following medium: 1 mmol EDTA + 215 mmol mannitol + 75 mmol sucrose + 0.1% BSA solution + 20 mmol HEPES (pH 7.2), followed by a double centrifugation in modes of 1,400g → 3 min. at 4°C (supernatant was removed) and 13000g → 10 min. The resulting secondary supernatant was removed for a respirometric analysis. The second part of the hippocampus was homogenized in PBS with pH 7.4 in a ratio of 1:7, centrifuged in the mode of 10000g → 5 min, and then the resulting supernatant was taken for ELISA.

### Respirometric analysis

In the work, the previously described approach to assessing mitochondria respirometric functions had been used. The study was performed according to the SEAHORSE protocol on a laboratory AKPM-01L respirometer (Alfa-Bassens, Russian Federation). During the analysis, the change in oxygen consumption in the medium containing native mitochondria, was assessed against the background of the injection of mitochondrial respiration disconnectors: oligomycin 1 µg/ml; 4-(trifluoromethoxy) phenyl) hydrazono) malononitrile (FCCP – 1 µM); rotenone – 1 µM; sodium azide – 20 mmol. Glucose (15

mmol) was used as an oxidation substrate in the process of the study of the intensity of anaerobic processes. At the same time, the following parameters characterizing the mitochondria respiratory function were calculated: ATP-generating capacity; maximum respiratory rate, respiratory capacity. The glycolysis intensity, glycolytic capacity, and glycolytic reserve were also determined [13].

### Enzyme-linked immunosorbent assay immunoassay

The standard ELISA reagents kits manufactured by Cloud clone (USA) was used. During the study, the changes in the following parameters were evaluated: β-amyloid concentration, apoptosis-inducing factor (AIF), endonuclease G (ENDOG) and ATP concentration. The analysis was carried out in accordance with the manufacturer's recommendations (a protocol analysis instruction was attached to each kit). The results measurement was carried out on a microplate reader Infinite F50 (Tecan, Austria).

### Methods of statistical analysis

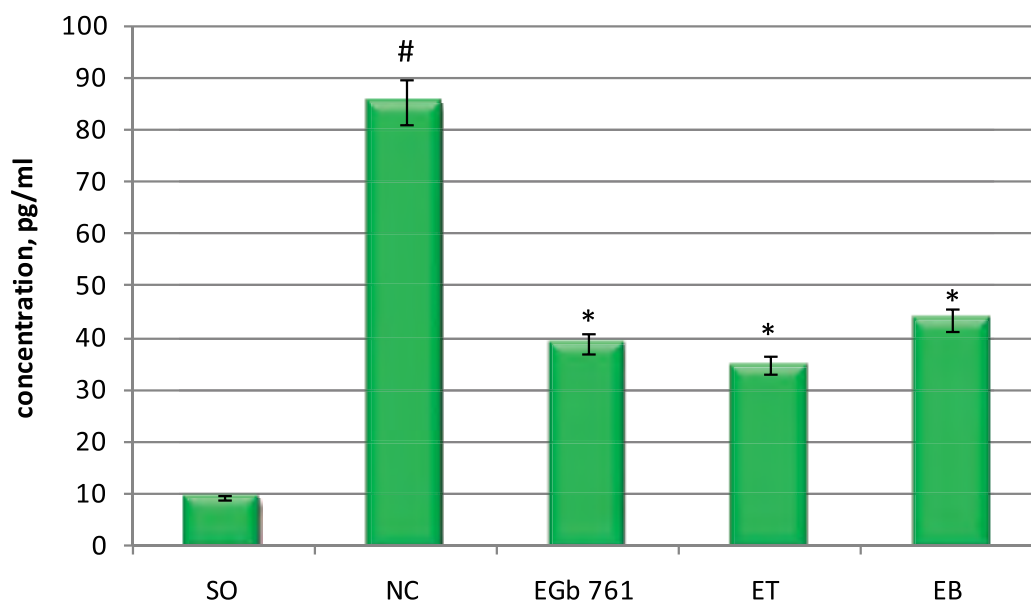
Statistical analysis of the obtained results was performed in the STATISTICA 6.0 software package (StatSoft, USA). The data were expressed as  $M \pm SEM$ . The comparison of means was carried out by the one-way analysis of the variance method with the post-hoc Newman-Keuls test.

**RESULTS****The influence of the test objects and the reference drug on the change of the  $\beta$ -amyloid concentration in the rats' hippocampus within the conditions of experimental acute brain hypometabolism**

During this part of the work it was found out that the  $\beta$ -amyloid concentration (Fig. 2) in the SO group was  $9.46 \pm 0.09$  pg/ml. At the same time, in the NC group of the animals within the conditions of the experimental

cerebral hypometabolism, an increase of the amyloid  $\beta$ -peptide concentration in comparison with the SO group of the animals was by 9 times ( $p < 0.0001$ ).

The use of EGb 761 reduced the content of cytotoxic  $\beta$ -amyloid in the rats' hippocampus in relation to the NC group of the animals by 54.4% ( $p < 0.0002$ ). Against the background of the test pumpkin and marigold extracts administration, the  $\beta$ -amyloid concentration decreased relative to the NC group of the animals by 54.4% ( $p < 0.0002$ ) and by 49% ( $p < 0.0002$ ), respectively (Fig. 2).



**Figure 2 – Effect of the test objects and the reference drug on the change of the  $\beta$ -amyloid concentration in the rats' hippocampus within the conditions of the experimental acute brain hypometabolism**

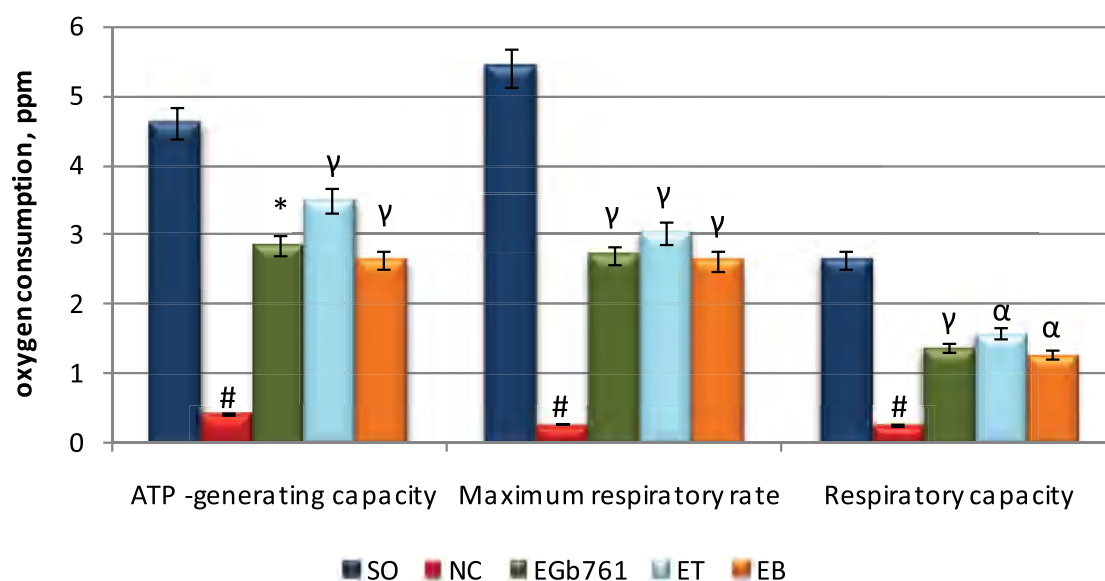
Note. SO – a sham-operated group of rats; NC – a negative control group of rats; ET – a group of rats treated with the test pumpkin extract; EB – a group of rats treated with the test marigold extract; EGb 761 – a group of rats treated with EGb 761; # – statistically significant relative to the SO animals group (Newman-Keuls test,  $p < 0.0001$ ); \* – statistically significant relative to the NC animals group (Newman-Keuls test,  $p < 0.0002$ ).

**The influence of the test objects and the reference drug on the change of the rats' hippocampal mitochondria respirometric function within the conditions of the experimental acute brain hypometabolism**

When assessing the effect of the test extracts and EGb 761 on the change of the hippocampal mitochondria respirometric function, it was found out that in the NC group within the conditions of the experimental brain hypometabolism compared with the SO group of rats, a decrease in the ATP-generating capacity was by

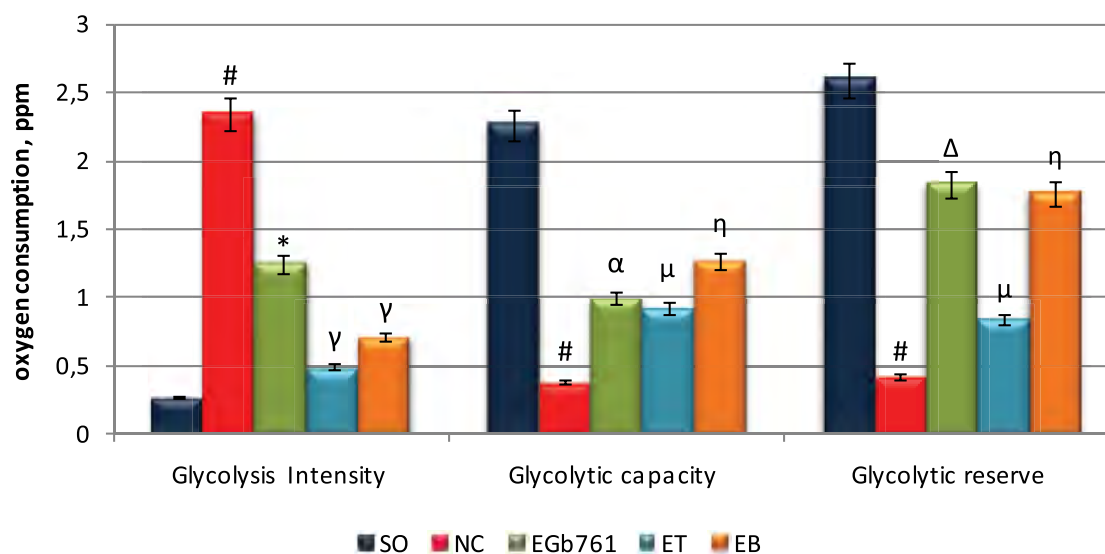
10.8 times ( $p < 0.0001$ ), the maximum respiratory rate was by 19.4 times ( $p < 0.0001$ ) and the respiratory capacity was by 9.9 times ( $p < 0.0001$ ) (Fig. 3).

In the animals deprived of pharmacological support, in relation to the SO group of rats, the glycolysis intensity increased by 8.5 times ( $p < 0.0001$ ), and the glycolytic capacity and glycolytic reserve decreased by 5.8 times ( $p < 0.0001$ ) and 6.1 times ( $p < 0.0001$ ), respectively (Fig. 4). As a result, the ATP concentration in the hippocampal supernatant of the NC group decreased by 2.2 times ( $p < 0.0001$ ) relative to the SO group (Fig. 5).



**Figure 3 – Effect of the test objects and the reference drug on the change of the mitochondrial respiration in the rats' hippocampus within the conditions of the experimental acute brain hypometabolism**

Note. SO – a sham-operated group of rats; NC – a negative control group of rats; ET – a group of rats treated with the test pumpkin extract; EB – a group of rats treated with the test marigold extract; EGb 761 – a group of rats treated with EGb 761; # – statistically significant relative to the SO group of animals (Newman-Keusle test,  $p < 0.0001$ ); statistically significant relative to the NC group of animals (Newman-Keusle test \* –  $p < 0.02$ ;  $\gamma$  –  $p < 0.001$ ;  $\alpha$  –  $p < 0.004$ ).



**Figure 4 – The effect of the test objects and the reference drug on the change of the anaerobic processes activity in the rats' hippocampus within the conditions of the experimental acute brain hypometabolism**

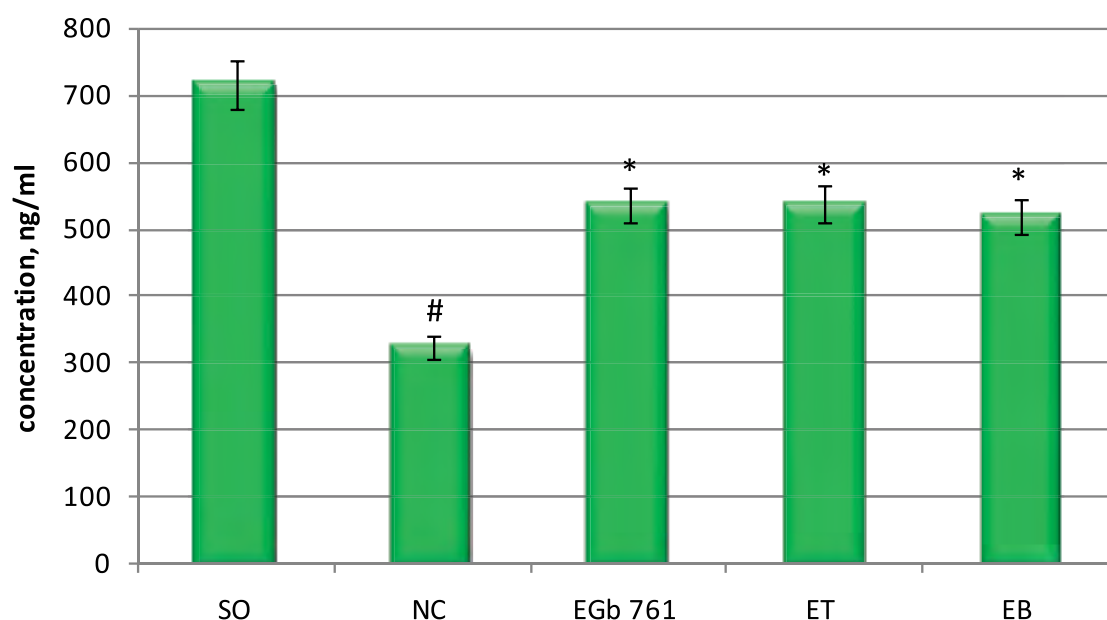
Note. SO – a sham-operated group of rats; NC – a negative control group of rats; ET – a group of rats treated with the test pumpkin extract; EB – a group of rats treated with the test marigold extract; EGb 761 – a group of rats treated with EGb 761; # – statistically significant relative to the SO animals' group (Newman-Keusle test,  $p < 0.0001$ ); \* – statistically significant relative to the NC group of animals (Newman-Keusle test, \* –  $p < 0.005$ ;  $\gamma$  –  $p < 0.002$ ;  $\alpha$  –  $p < 0.007$ ;  $\Delta$  –  $p < 0.03$ ;  $\mu$  –  $p < 0.01$ ;  $\eta$  –  $p < 0.004$ ).



Against the background of EGb 761 administration in rats, an increase (relative to the NC group of animals) of ATP-generating capacity (Fig. 3) by 6.7 times ( $p < 0.002$ ), the maximum respiratory rate by 9.7 times ( $p < 0.001$ ) and respiratory capacity – by 5.1 times ( $p < 0.001$ ) was noted. In the rats treated with EGb 761, there was also an increase in glycolytic capacity and glycolytic reserve (Fig. 4) in comparison with the same parameters of the NC animals' group by 2.6 times ( $p < 0.007$ ) and by 4.3 times ( $p < 0.03$ ), respectively, while the intensity of glycolysis decreased by 1.9 times ( $p < 0.005$ ). In addition, the concentration of ATP in the animals treated with EGb 761 was 65.7% ( $p < 0.002$ ) higher than that in the rats without any pharmacological correction.

In the animals treated with the pumpkin extract, rel-

ative to the NC rats' group, an increase in ATP-generating capacity was by 8.2 times ( $p < 0.001$ ); the maximum level of respiration was by 10.9 times ( $p < 0.001$ ) and respiratory capacity was by 5.9 times ( $p < 0.004$ ) (Fig. 3). At the same time, the intensity of glycolysis in the animals that were administrated pumpkin extract, was 4.7 times ( $p < 0.002$ ) lower than that in the NC group of rats, while the glycolytic capacity and glycolytic reserve (Fig. 4) in the animals treated with the test pumpkin extract, increased in comparison with the rats deprived of pharmacological support by 2.4 times ( $p < 0.01$ ) and twice ( $p < 0.004$ ), respectively. The administration of the pumpkin extract in the animals contributed to an increase of ATP content (Fig. 5) in the hippocampus by 66.2% ( $p < 0.002$ ) in relation to the NC group of rats.



**Figure – 5. The effect of the test-objects and the reference drug on the change of the ATP concentration in the rats' hippocampus within the conditions of the experimental acute brain hypometabolism**

Note. SO – a sham-operated group of rats; NC – a negative control group of rats; ET – a group of rats treated with the test pumpkin extract; EB – a group of rats treated with the test marigold extract; EGb 761 – a group of rats treated with EGb 761; # – statistically significant relative to the SO group (Newman-Keusle test,  $p < 0.0001$ ); \* – statistically significant relative to the NC group (Newman-Keusle test,  $p < 0.002$ ).

When the marigold extract was administrated in the animals, an increase (relative to the NC rats' group) of ATP-generating capacity, the maximum respiratory rate and respiratory capacity (Fig. 3) by 6.2 times ( $p < 0.001$ ); by 9.4 ( $p < 0.001$ ) and 4.7 times ( $p < 0.004$ ), respectively, were noted. In addition, the intensity of glycolysis in the animals treated with marigold extract was 3.3 times lower ( $p < 0.002$ ) than that in the NC group of rats, while the glycolytic capacity and glycolytic reserve (Fig. 4) increased in the animals treated with marigold extract compared with the rats deprived of pharmacological support by 3.3 ( $p < 0.01$ ) and 4.2 times ( $p < 0.004$ ), respectively. As a result, the ATP concentration in the hippocampus of the animals treated with the marigold extract

was 60.7% ( $p < 0.002$ ) higher than that of the rats of the NC group (Fig. 5).

#### The influence of the test objects and the reference drug on the change of the internal apoptosis pathway activity within the conditions of experimental acute brain hypometabolism

In the course of this part of the study it was found out that the concentration of AIF and ENDOG in the animals of the NC group exceeded the similar values of the SO rats' group by 3 ( $p < 0.0001$ ) and 21.7 times ( $p < 0.002$ ), respectively (Table 1). At the same time, the animals treated with EGb 761 showed a decrease in AIF concentration by 36.8% ( $p < 0.002$ ) and ENDOG by 2.7 times

( $p < 0.0001$ ) compared with the group of rats deprived of pharmacological support (Table 1).

Against the background of using the test pumpkin extract in the animals, the contents of AIF and ENDOG in the hippocampus decreased (relative to the NC group of rats) by 33% ( $p < 0.002$ ) and 3.6 times ( $p < 0.002$ ), respectively. A decrease in the concentration of proapop-

totic markers of the internal apoptosis pathway was also observed while using the marigold extract. So, in the animals that were administrated the test marigold extract, the concentration of AIF and ENDOG was 38.3% ( $p < 0.002$ ) and 4.4 times ( $p < 0.002$ ), respectively, lower than the similar parameters of the NC rats' group (Table 1).

**Table 1 – The influence of the test objects and the reference drug on the change of the internal apoptosis pathway activity within the conditions of experimental acute brain hypometabolism**

Group	SO	NC	EGb 761	ET	EB
AIF, ng/ml	6.16±0.413	18.69±0.04#	11.81±0.043*	12.61±0.426*	11.54±0.446*
ENDOG, ng/ml	72.03±1.069	1564.42±63.59#	578.41±60.45*	433.09±42.41*	352.35±10.796*

Note. AIF – apoptosis-inducing factor; ENDOG – endonuclease G; SO – a sham-operated group of rats; NC – a negative control group of rats; ET – a group of rats treated with the test pumpkin extract; EB – a group of rats treated with the test marigold extract; EGb 761 – a group of rats treated with EGb 761; # – statistically significant relative to the SO animals' group (Newman-Keuls test,  $p < 0.0001$ ); \* – statistically significant relative to the NC group of animals (Newman-Keuls test,  $p < 0.0002$ ).

## DISCUSSION

Currently, it has been established that the cerebral hypometabolism is one of the leading factors for the AD development. An experimental study by *Riha et al., 2008* provides the data that hypometabolism of the posterior cingulate cortex contributes to the deposition of toxic amyloid  $\beta$ -peptide in this brain structure, resulting in characteristic clinical manifestations of AD in animals: decreased cognitive functions, activation of lipid peroxidation and decreased synaptic transmission [14]. This study was completed by *Scheltens et al., 2018*, who proved that in addition to impaired memory and associated brain functions, a decrease in metabolic processes contributes to the development of neurological symptoms, motor and sensory areflexion [15]. Brain hypometabolism is largely associated with impairment of the cellular bioenergetic processes, depending on the functional activity of mitochondria [16]. *In vitro* and *in vivo* approaches to the carried out studies have shown that a decrease in the mitochondrial function is observed before the development of deep structural changes in the brain, characteristic to Alzheimer's pathology, and include a decrease of mitochondrial respiration, expression and activity of metabolic enzymes, an increase oxidative stress and expression of a proapoptotic signal [17]. Hereby, the activation of the apoptotic cascade occurs mainly due to the increase of the intensity of internal apoptosis pathway reactions (depending on mitochondria) with an increased apoptosis-inducing factor releasing and endonuclease G activation [18]. The induction of apoptosis enhances the cytotoxicity of  $\beta$ -amyloid, which accelerates the death of neurons and, accordingly, worsens the course of disease [19]. Previously, it was established that about 90% AD patients have hippocampal hypometabolism, and the clinical symptoms of AD varied greatly: from minor neurological disorders to diffuse cognitive deficit, accompanied by motor fluctuations and loss of sensory perception [20].

In addition, people with cerebral hypometabolism are, as a rule,  $\beta$ -amyloid-positive patients whose prognosis is most unfavorable [21].

In this regard, it can be assumed that the correction of hippocampal hypometabolism can be a promising approach to preventive pharmacotherapy of AD, which was partially confirmed by *Villain et al., 2008* [22]. However, in spite of the probable expediency of early correction of metabolic disorders, provided they are diagnosed in a timely manner, currently existing AD treatment methods are aimed at reducing the cytotoxicity of  $\beta$ -amyloid, and the spectrum of compounds which can restore the metabolic activity of cells, is quite limited [23]. In addition, the developed drugs of targeted AD correction often have undesirable toxicological parameters. An example of this agent is semagacestat, a  $\gamma$ -secretase inhibitor, a potentially effective drug for the treatment of AD, which reduces the toxicity of  $\beta$ -amyloid, but has a high oncogenic potential, and therefore its further study has been stopped [24].

In this regard, a study to evaluate the effect of pumpkin (*Cucurbita pepo* L.) and marigold extracts (*Tagetes patula* L.) on the functional activity of hippocampal mitochondria within the conditions of the experimental acute brain hypometabolism was conducted. During the study, it was found out that the prophylactic administration of the tests pumpkin and marigold extracts contributed to the restoration of the mitochondrial function and a decrease of the anaerobic processes intensity equally with the reference drug – a standardized *Ginkgo biloba* extract (EGb 761).

Moreover, in the groups of the rats treated with marigold, pumpkin extracts and EGb 761, there was an increase of ATP concentration in the hippocampal supernatant compared to the same indicator in the group of the rats deprived of pharmacological support by 65.7% ( $p < 0.002$ ); 66.2% ( $p < 0.002$ ); and 60.7% ( $p < 0.002$ ), respectively. It may indicate normalization of bioenergetic



processes in the hippocampus within the conditions of acute cerebral hypometabolism when using the test extracts and the reference drug [25].

In addition, the restoration of the optimal ATP synthesis can prevent the destabilization of mitochondrial membranes, which prevents their decomposition and release of AIF, thereby inhibiting internal apoptotic cascade reactions [26]. That was also established during the study. So, when the animals were treated with pumpkin and marigold extracts, a decrease of the AIF concentration (by 33% ( $p < 0.002$ ) and 38.3% ( $p < 0.002$ ), respectively), and ENDOG (by 3.6 times ( $p < 0.002$ ) and by 4.4 times ( $p < 0.002$ ), respectively), was noted.

Against the background of the test extracts and EGb 761, a decrease in the concentration of amyloid  $\beta$ -peptide in the rats' hippocampus by 54.4% ( $p < 0.0002$ ); by 54.4% ( $p < 0.0002$ ) and by 49% ( $p < 0.0002$ ), respectively,

was noted. As a whole with a decrease of apoptosis intensity, it can prevent neuronal destruction [27].

### CONCLUSION

On the basis of the obtained data, it can be assumed that the prophylactic administration of pumpkin extract and marigold extract helps to normalize bioenergetic processes, decrease the concentration of proapoptotic markers and  $\beta$ -amyloid in the rats' hippocampus under acute hypometabolism caused by intrahippocampal administration of a 3M sodium azide solution.

At the same time, the test extracts showed an equivalent therapeutic efficacy with the reference drug – a standardized extract of *Ginkgo biloba* (EGb 761), which makes these extracts promising objects for a further study in order to create drugs for targeted correction of AD in the early hypometabolic stage of the disease.

### FINANCIAL SUPPORT AND SPONSORSHIP

This study did not have any financial support from outside organizations.

### AUTHOR CONTRIBUTIONS

All authors had equally contributed to the research work.

### CONFLICTS OF INTEREST

The authors and peer reviewers of this paper report no conflicts of interest.

### ACKNOWLEDGMENTS

The authors are grateful Serebryanaya F.K. for help in the authenticity of plant raw materials determining.

### REFERENCES

- Gao C, Chang P, Yang L. Neuroprotective effects of hydrogen sulfide on sodium azide-induced oxidative stress in PC12 cells. *Int J Mol Med*. 2018; 41(1):242–250. doi:10.3892/ijmm.2017.3227
- Takahashi RH, Nagao T, Gouras GK. Plaque formation and the intraneuronal accumulation of  $\beta$ -amyloid in Alzheimer's disease. *Pathol Int*. 2017; 67:185–193. doi: 10.1111/pin.12520.
- Chen GF, Xu TH, Yan Y. Amyloid beta: structure, biology and structure-based therapeutic development. *Acta Pharmacol Sin*. 2017;38(9): 1205–1235. doi:10.1038/aps.2017.28
- Lesné SE, Sherman MA, Grant M. Brain amyloid- $\beta$  oligomers in ageing and Alzheimer's disease. *Brain*. 2013; 136, Part 5: 1383–1398. doi:10.1093/brain/awt062
- Mattson MP. Pathways towards and away from Alzheimer's disease. *Nature*. 2004; 430(7000): 631–639. doi:10.1038/nature02621.
- Del Prete D, Suski JM, Oulès B. Localization and Processing of the Amyloid- $\beta$  Protein Precursor in Mitochondria-Associated Membranes. *J Alzheimers Dis*. 2017;55(4): 1549–1570. doi:10.3233/JAD-160953
- Lustbader JW, Cirilli M, Lin C, Xu HW, Takuma K, Wang N, Caspersen C, Chen X, Pollak S, Chaney M. Abad directly links a beta to mitochondrial toxicity in Alzheimer's disease. *Science*. 2004;304:448–452. doi: 10.1126/science.1091230.
- Chen X, Yan SD. Mitochondrial abeta: A potential cause of metabolic dysfunction in Alzheimer's disease. *IUBMB Life*. 2006;58:686–694. doi: 10.1080/15216540601047767.
- De Strooper B, Iwatsubo T, Wolfe MS. Presenilins and  $\gamma$ -secretase: structure, function, and role in Alzheimer Disease. *Cold Spring Harb Perspect Med*. 2012;2(1):a006304. doi:10.1101/cshperspect.a006304
- Zhao Y, Zhang Y, Pan F. The effects of EGb761 on lipopolysaccharide-induced depressive-like behaviour in C57BL/6J mice. *Cent Eur J Immunol*. 2015;40(1): 11–17. doi:10.5114/ceji.2015.49427
- Gordon RYA, Kapralova MV, Goduhin OV, Arhipov VI. Osobennosti narushenij pamyati u krys posle povrezhdeniya polya SA3 dorsal'nogo gippokampa kainovoj kislotoj [Features of memory impairment in rats after damage to the CA3 field of the dorsal hippocampus with kainic acid] *Bulletin of Experimental Biology and Medicine*. 2013;155(6): 771–775 Russian
- Brouillet E, Hyman BT, Jenkins BG, Henshaw DR, Schulz JB, Sodhi P, Rosen BR, Beal MF. Systemic or local administration of azide produces striatal lesions by an energy impairment-induced excitotoxic mechanism. *Experimental Neurology*. 1994;129:175–182.
- Voronkov A.V., Pozdnyakov D.I., Nigaryan S.A., Khouri E.I., Miroshnichenko K.A., Sosnovskaya A.V., Olokhova E.A. Evaluation of the mitochondria respirometric function in the conditions of pathologies of various geneses. *Pharmacy & Pharmacology*. 2019;7(1):20–31. doi:10.19163/2307-9266-2019-7-1-20-31
- Riha PD, Rojas JC, Colorado RA, Gonzalez-Lima F. Animal model of posterior cingulate cortex hypometabolism implicated in amnesic MCI and AD. *Neurobiol Learn Mem*. 2008;90(1):112–124. doi:10.1016/j.nlm.2008.01.011

15. Scheltens NME, van der Weijden K, Adriaanse SM. Hypometabolism of the posterior cingulate cortex is not restricted to Alzheimer's disease. *Neuroimage Clin.* 2018;19: 625–632. doi:10.1016/j.nicl.2018.05.024
16. Nicholson RM, Kusne Y, Nowak LA, LaFerla FM, Reiman EM, Valla J. Regional cerebral glucose uptake in the 3xTG model of Alzheimer's disease highlights common regional vulnerability across AD mouse models. *Brain Res.* 2010; 1347:179–185. doi:10.1016/j.brainres.2010.05.084
17. Chou JL, Shenoy DV, Thomas N, Choudhary PK, Laferla FM, Goodman SR. Early dysregulation of the mitochondrial proteome in a mouse model of Alzheimer's disease. *Journal of Proteomics.* 2011;74(4): 466–479. doi: 10.1016 / j.jprot.2010.12.012.
18. Li Z, Chen X, Lu W. Anti-Oxidative Stress Activity Is Essential for *Amanita caesarea* Mediated Neuroprotection on Glutamate-Induced Apoptotic HT22 Cells and an Alzheimer's Disease Mouse Model. *Int J Mol Sci.* 2017;18(8): 1623. doi:10.3390/ijms18081623
19. Obulesu, M, Jhansi Lakshmi M. Apoptosis in Alzheimer's disease: an understanding of the physiology, pathology and therapeutic avenues. *Neurochemical research.* 2014;39(12): 2301–2312.
20. Chételat G, Ossenkoppele R, Villemagne VL. Atrophy, hypometabolism and clinical trajectories in patients with amyloid-negative Alzheimer's disease. *Brain.* 2016; 139, Part 9: 2528–2539. doi:10.1093/brain/aww159
21. Sperling R, Mormino E, Johnson K. The evolution of pre-clinical Alzheimer's disease: implications for prevention trials. *Neuron.* 2014; 84(3): 608–622. doi:10.1016/j.neuron.2014.10.038
22. Villain N, Desgranges B, Viader F. Relationships between hippocampal atrophy, white matter disruption, and gray matter hypometabolism in Alzheimer's disease. *J Neurosci.* 2008; 28(24): 6174–6181. doi:10.1523/JNEUROSCI.1392-08.2008
23. Cummings J, Aisen PS, DuBois B. Drug development in Alzheimer's disease: the path to 2025. *Alzheimers Res Ther.* 2016;8: 39–51 doi:10.1186/s13195-016-0207-9.
24. Henley DB, Sundell KL, Sethuraman G, Dowsett SA, May PC. Safety profile of semagacestat, a gammasecretase inhibitor: IDENTITY trial findings. *Curr Med Res Opin.* 2014; 10: 2021–2032. doi: 10.1185/03007995.2014.939167.
25. Frenguelli BG. The Purine Salvage Pathway and the Restoration of Cerebral ATP: Implications for Brain Slice Physiology and Brain Injury. *Neurochem Res.* 2019;44(3):661–675. doi:10.1007/s11064-017-2386-6
26. Farina B, Di Sorbo G, Chambery A. Structural and biochemical insights of CypA and AIF interaction. *Sci Rep.* 2017;7(1):1138. doi: 10.1038/s41598-017-01337-8
27. Masters CL, Selkoe DJ. Biochemistry of amyloid  $\beta$ -protein and amyloid deposits in Alzheimer disease. *Cold Spring Harb Perspect Med.* 2012;2(6): a006262. doi:10.1101/cshperspect.a006262

## AUTHORS

**Andrey V. Voronkov** – Doctor of Science (Med.), professor, head of the department of pharmacology with a course of clinical pharmacology. Pyatigorsk medical-pharmaceutical Institute. ORCID ID:0000-0001-6638-6223. E-mail: prohor77@mail.ru

**Dmitry I. Pozdnyakov** – Candidate of I Sciences (Pharm.), Senior Lecturer of the Department of Pharmacology with a course of clinical pharmacology. Pyatigorsk medical-pharmaceutical Institute. ORCID ID:0000-0003-0889-7855. E-mail: pozdniackow.dmitry@yandex.ru

**Similya L. Adzhiakhmetova** – PhD, lecturer of the Department of Organic Chemistry of the Pyatigorsk Medical Pharmaceutical. E-mail: similla503@mail.ru

**Nadezhda M. Chervonnaya** – PhD, lecturer of the

Department of Organic Chemistry of the Pyatigorsk Medical Pharmaceutical Institute. E-mail: nadezhda.chervonnaya@yandex.ru

**Kirill A. Miroshnichenko** – PhD-student of the Department of Pharmacology with a course of clinical pharmacology. Pyatigorsk medical-pharmaceutical Institute. E-mail: K220436@yandex.ru

**Anastasia V. Sosnovskaya** – 5th year student of the Pharmaceutical department. Pyatigorsk medical-pharmaceutical Institute. ORCID ID: 0000-0002-5037-994X. E-mail: 88misi88@yandex.ru

**Elizaveta I. Shereshkova** – 5th year student of the Pharmaceutical department. Pyatigorsk medical-pharmaceutical Institute. ORCID ID: 0000-0002-7854-5401. E-mail: elizaveta.shereshkova@yandex.ru