



COLLAGENASES IN MEDICAL PRACTICE: MODERN COLLAGENASE-BASED PREPARATIONS AND PROSPECTS FOR THEIR IMPROVEMENT

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The aim of this study was to assess the coverage of studies of collagenolytic enzymes (collagenases) in the sphere of their use in wounds and scars treatment, the resource of their production and the range of collagenase products to identify the areas for their improvement.

Materials and methods. The information from retrieval and library databases (eLIBRARY, PubMed, Scopus, ScholarGoogle, ResearchGate), patent information databases (freepatent.ru, ntpo.com, fips.ru), the State register of medicines (GRSL) and the State register of medical devices, as well as technical information provided by manufacturers of medicines and medical devices, was used in the article.

Results. The analysis of the information database has shown that collagenolytic enzymes are effective proteolytic complexes because of their ability to provide the breakdown of collagen, the main component of wounds and scars.

Hepatopancreas crustaceans is currently one of the available raw resources of collagenases in Russia. It is noted that proteolytic enzymes from the *Paralithodes camtschatica* hepatopancreas are characterized by a broad specificity: they are able to hydrolyze both native collagen and other protein substrates. There are data confirming the capability of collagenases to accelerate the process of reparation in addition to wound cleansing from a necrosis. The results of clinical studies of collagenases anti-scar properties, indicate the effectiveness of their use for the skin scar correction. The content analysis has shown that there is a small amount of collagenase-based products in the Russian pharmaceutical market: lyophilized powder for preparation of the injection solution "Collalysin", recommended for scars treatment; a medical dressing "Digestol" with collagenase, recommended for wounds and necrotic lesions treatment; "Fermencol" (gel and powder), the "Karipain plus" gel for scars treatment. Drugs are represented by only powder lyophilisate "Collalysin".

Conclusion. The development of gel compositions (Aerosil-based oleogels) and atraumatic dressings with collagenase from *Paralithodes camtschatica* hepatopancreas as the most affordable raw materials can be considered problem number one of practical pharmacy at present. This provides for the creation of the dosage forms, improved in terms of stability and efficiency, as well as ease of use.

Keywords: collagenolytic enzymes, *Paralithodes camtschatica* collagenase, scars, wounds, preparations, gels

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КОЛЛАГЕНАЗЫ В МЕДИЦИНСКОЙ ПРАКТИКЕ: СОВРЕМЕННЫЕ СРЕДСТВА НА ОСНОВЕ КОЛЛАГЕНАЗЫ И ПЕРСПЕКТИВЫ ИХ СОВЕРШЕНСТВОВАНИЯ

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Цель. Оценка состояния изученности применения коллагенолитических ферментов (коллагеназ) в терапии ран и рубцов, источников их получения, ассортимента средств с коллагеназой для выявления направлений их совершенствования.

Материалы и методы. В работе использованы информационно-поисковые и библиотечные базы данных (eLIBRARY, PubMed, Scopus, ScholarGoogle, ResearchGate), патентно-информационные базы (freepatent.ru, ntpo.com, fips.ru), Государственный реестр лекарственных средств и Государственный реестр медицинских изделий, а также техническая информация, представленная производителями лекарственных средств и медицинских изделий.

Результаты. Анализ информационных источников показал, что коллагенолитические ферменты являются эффективными протеолитическими комплексами, т.к. обладают способностью обеспечивать расщепление коллагена, являющегося главным компонентом ран и рубцов. Одним из доступных сырьевых источников коллагеназ в России в настоящее время является гепатопанкреас ракообразных. Отмечается, что для протеолитических ферментов из гепатопанкреаса камчатского краба характерна широкая специфичность: они способны гидролизовать как нативный коллаген, так и другие белковые субстраты. Имеются данные исследований, подтверждающие, что коллагеназы наряду с очисткой раны от некроза способны ускорять процесс ее репарации. Результаты клинических исследований противорубцовых свойств коллагеназ свидетельствуют об эффективности их использования для коррекции рубцовых изменений кожи. Контент-анализ показал, что на российском фармацевтическом рынке присутствует незначительное количество средств на основе коллагеназ – лиофилизированный порошок для приготовления раствора для инъекций («Коллализин»), рекомендуемый для лечения рубцов; медицинская повязка с коллагеназой, рекомендуемые для лечения ран и некротических поражений «Дигестол»; гель и порошок «Ферменкол», гель «Карипаин плюс» для лечения рубцов. Лекарственные препараты представлены только порошком-лиофилизатом «Коллализин».

Заключение. Разработка состава гелей (олеогелей на основе аэросила) и атравматических салфеток с коллагеназой из гепатопанкреаса краба как наиболее доступного сырья можно считать актуальной задачей практической фармации. Это предусматривает создание лекарственных форм, улучшенных с позиций стабильности и эффективности, а также удобства использования.

Ключевые слова: коллагенолитические ферменты, коллагеназа камчатского краба, рубцы, раны, препараты, гели

INTRODUCTION

The problem of effective and rapid healing of wounds resulting from injuries of different origins, remains one of the most relevant in modern medical practice. Currently, the effectiveness of wound healing is determined not only by the term, but also by the aesthetic result achieved. In addition, special attention is paid to the treatment convenience to combine the optimal therapeutic effect and a patient's life quality [1].

In medical practice, preparations of proteolytic enzymes are used to accelerate the granulation process and reduce the time of wound healing by the ability

to cleanse wounds from necrotic tissues and exudate. Preparations of proteolytic enzymes (collagenase, hyaluronidase) are also used for scars medical correction [2, 3].

The use of enzyme preparations in the composition of external therapy, has some peculiarities caused mainly by their instability, which makes it relevant to improve the existing dosage forms both in terms of stability and efficiency, and ease of use. Collagenases are one of the most effective proteolytic enzymes due to their ability to provide collagen, the main component of wounds and scars. Thus, studying the problems related

to collagenolytic enzymes raw resources, mechanisms of their action, clinical efficacy of collagenases in wounds and scars, as well as the range and features of the use of collagenase-based products in medical practice, are relevant.

THE AIM of the review is to assess the research of collagenolytic enzymes (collagenases) use in the treatment of wounds and scars, the sources of their production, the range of collagenase products to identify the areas for their improvement.

MATERIALS AND METHODS

The information from retrieval and library databases (eLIBRARY, PubMed, Scopus, ScholarGoogle, ResearchGate for the time interval from 2000 to 2019), patent information databases (freepatent.ru, ntpo.com, fips.ru), reference literature, the State Register of Medicinal Remedies (SRMR) and the State Register of Medical Products, (SRMP) as well as technical information provided by manufacturers of medicines and medical products, the sites dedicated to cosmetic products "Fermencol" and "Karipain", is used in the article. The depth of the patent search was 30 years. The keywords in the search process are: collagenase, collagenolytic enzymes, enzyme immobilization, wounds, treatment, scars.

RESULTS AND DISCUSSION

The role of enzyme preparations in the correction of pathological wound healing

Wound healing is a complex biological process that consists of overlapping phases: inflammation, proliferation, and remodeling. Wound healing is a regulated process in which several cell types (keratinocytes, fibroblasts, endothelial cells, macrophages and platelets) and a network of signaling molecules (cytokines, chemokines and growth factors) are in effect. In its course, the disorders which can lead to hard-to-heal chronic wounds or scars are possible. The most common factors impeding normal healing are diabetes, venous disease, old age, peripheral neuropathy, impaired microflora and malnutrition. A wound is considered chronic when it does not show a tendency to repair for more than 4 weeks. So, according to the data, full and complete wound closure, is achieved in only 25-50% of cases of chronic or hard-to-heal wounds after 20 weeks' treatment, especially in the case of venous and diabetic ulcers. Chronic hard-to-heal wounds are often characterized by a number of microbiological (increased microbial contamination, including the presence of biofilms), biochemical and cellular pathologies that prevent or slow down the process of healing. Unlike acute wounds, chronic wounds are not completed by the process of remodeling, but are considered as a process stopped at the stage of inflammation or proliferation [1-4].

The presence of cellular detritus, necrotic tissues and bacterial toxins leads to the prolongation of inflammation and increased production of cytokines by mac-

rophages and neutrophils, resulting in the activation of macrophages and fibroblasts. An excessive activity of tissue proteases is observed [3].

The so-called "wound bed preparation" technology is used to remove the necrotic component to transfer the wound from a chronic state to an acute one. Traditionally, to remove necrotic, damaged or infected tissues, different methods are used: surgical, autolytic, enzymatic, chemical and physical. Wound cleansing helps to reduce the presence of inflammatory cytokines and metalloproteinases, which are produced during chronic wounds inflammation [4].

One of the problems of pathological wound healing is the scars (keloid and hypertrophic) formation, which occurs when the regulation by fibroblasts and keratinocytes balance of collagen synthesis-breakdown process disrupts, and the disturbance of the collagen remodeling process takes place. A stimulating effect on fibroblasts is provided by a chronic inflammation of the scar tissue, a long-term wound healing, secondary infections and epithelialization disruption [1].

Enzymatic wound cleansing is an effective and selective method, often used in combination with other methods of treatment, e.g., in combination with moisturizing dressings. In chronic wounds, it is necessary not only for cleansing (removal of necrotic tissue), but also for the migration of cells involved in epithelialization, as well as the elimination of inflammation. Unlike acid preparations used for treatment of wounds with a high content of necrotic tissues, e.g., the ointment containing salicylic acid 40%, proteases have no effect on intact tissues [2, 3].

Collagen is the most stable protein of tissue detritus. In this case, the wounds containing collagen fibers, hardly give way to enzymatic cleansing by means of other proteolytic enzymes: trypsin, chymopsin, papain, terilitin, streptokinase, etc. [2]. Collagenase preparations are successfully used to treat wounds with massive purulonecrotic discharge, trophic ulcers, frostbite, burns, scars. Proteolytic enzymes with collagenolytic activity are the most effective for the wounds' treatment and elimination of scarring [5-11].

Characteristics of collagenases used in medical practice: sources of production, mechanism of action, data of pharmacological and clinical studies

Collagenase is a specific proteolytic enzyme that breaks down peptide bonds in natural collagen, the main structural element of a connective tissue. According to the active ingredient resource, collagenase preparations are biological agents, because their industrial raw materials are microorganisms' cultures or animals' digestive glands.

The analysis of the commercial drugs, veterinary and medical products, cosmetics composition with collagenases showed, that modern products contain enzyme

complexes, the source of which are the *Clostridium* family bacteria or digestive tract glands (hepatopancreas) of the *Paralithodes camtschaticus* [12–15].

Collagenases derived from *Clostridium histolyticum*, the most commonly used drug-derived microbial proteases, are single-stranded proteins with masses ranging from 68 kDa to 130 kDa. Thus, the “Santil” ointment contains two collagenases (collagenase G, ~114 kDa, and collagenase H, ~110 kDa), nonspecific neutral metalloproteinase (~35 kDa), a small amount of cysteine proteases (clostripain ~58 kDa). The disadvantages of microbial collagenase preparations are: a relatively low activity, difficulties in cultivating producers, as well as potential allergenicity, which may be caused by the usage of pathogenic microorganisms-producers [4, 11, 12].

In Russia, hepatopancreas *Paralithodes* of crustaceans is widely used as a source of collagenase, which is considered as an affordable, cheap and non-toxic raw material, being a waste of commercial crab processing. The research of this group of proteases is devoted to both the production of stable and highly active enzyme complexes and the development of effective drugs [13–19].

Hepatopancreas crustaceans produces a number of digestive enzymes that hydrolyze different classes of biopolymers: collagenolytic (serine, trypsin-like) proteases, collagenases, phosphatases, phosphodiesterases, elastases, RNases, DNases, etc. It has been proved that proteolytic enzymes of *Paralithodes camtschaticus*, possess a broad specificity. They disintegrate both native collagen and other protein substrates – casein, gelatin, fibrinogen and serum albumin, which, in many respects, leads to the high efficiency of these complexes [14, 16, 20]. The enzyme complexes obtained from the *Paralithodes camtschaticus* hepatopancreas, vary according to the degree of purity, activity and composition. Thus, the sum of collagenolytic proteases of crab hepatopancreas, which is a mixture of nine proteins with a molecular weight of 23–36 kDa, was isolated. In the preparations “Collalitin” and “Collagenase KK”, collagenolytic proteases of *Paralithodes camtschaticus* hepatopancreas are represented in the form of three isoenzymes, the molecular weight of which is in the range from 18 to 27 kDa [12, 15–20].

The mechanism of a therapeutic action of collagenases is based on the ability to convert native insoluble collagen into a soluble form by hydrolysis of peptide bonds. True collagenases, particularly of microbial and animal origin (the metalloproteases class), cleave the triple helix of collagen at one specific point, forming large soluble fragments, further destruction of which is relatively slow. It has been established that purified clostridial collagenase, as well as the complex of enzymes present in the “Santil” preparation, hydrolyze native and denatured collagen, as well as collagen-associated proteins of the intercellular matrix to peptides. Serine proteases (collagenolytic proteases) obtained from the

gastrointestinal tract of fishes and invertebrates, break down the three polypeptide chains of tropocollagen, and the resulting peptides are further hydrolyzed to amino acids [2, 11, 12, 20–26].

It is noted that in the wound, proteolytic enzymes contribute to the exudate colliquation, facilitating the access for antiseptic and antibiotic drugs to the bacterial cell, enhancing the effect of antibacterial therapy. On the model of an infected rats’ burn it was established that the collagenase treatment of the wound formation from day 5 from the beginning of the process reduced a bacterial load from 108 to 105 or fewer bacteria per gram of tissue. The level of bacterial load had a beneficial effect on normal wound healing, which contributed to the reparation acceleration. The authors suggest that collagenases can be safely used without concomitant local antimicrobials in chronically infected wounds due to their antimicrobial properties and effectiveness of wound healing [25, 26].

In the studies it has been established, that in addition to cleansing the wound of necrotic tissues, collagenases directly affect the reparation process. Thus, collagenase isolated from *C. histolyticum*, was found to enhance migration and proliferation of keratinocytes, endothelial cells and fibroblasts. The study of the effect of bacterial collagenase preparations on the model of full-layer wound in Yucatan pigs showed, that the daily treatment with the enzyme from the first day of the wound formation, made it possible to achieve purification, exudation, as well as effective angiogenesis and epithelization and, as a result, effective wound healing in a shorter time [28].

The reparative effect of proteolytic complexes obtained from crustaceans, has also been revealed. The study of moricrasa (a lipophilic base ointment with *Paralithodes camtschaticus* collagenase) reparative properties on the spontaneous purulent-ulcer injuries in rats, showed a complete wound healing after 6–10 days, when applied daily. It has also been noted that the further application of the ointment, contributed to the resumption of the wool cover at the wound location [29].

The study of reparative properties of oleogel with *Paralithodes camtschaticus* collagenase on the rats’ burn skin model showed, that the test ointment, applied every day beginning from the 3rd day after the infection and formation of spontaneous purulent-necrotic wounds, activated the processes of epithelization and proliferation in the damaged tissues, which significantly reduced the healing time – there was complete epithelization of the burn surface, desquamation of the scab on the 10th day, in comparison with the control group, where incomplete epithelization was observed [30].

In the experiments on the animals (rats) it was demonstrated, that the wounds treatment with chitosan-modified textile with immobilized collagenase of *Paralithodes camtschaticus*, significantly reduced the necrotic tissue to 3 days (14 days in the control) and the

purulent wounds healing time reduced to 12 days (27 days in the control) [18].

Attempts to research the mechanism of microbial collagenases influence on the wound healing effectiveness were carried out. *In vitro* research has established, that the intercellular matrix enzymolysis with bacterial collagenase and the "Santil" ointment, containing collagenases, releases peptides that activate cellular migration, proliferative and angiogenic processes in trauma and promote wound healing. *In vitro* research has also revealed that collagen hydrolysis fragments and collagen-associated peptides obtained as a result of endothelial collagenase hydrolysis of dermal capillaries and human fibroblasts, increase cell proliferation and promote angiogenesis. On the model of full-layer long-term non-healing wounds of the mice, it was demonstrated that collagenase of "Santil" ointment, as well as peptides obtained from extracellular matrix, increase wound reepithelization by 60–100% compared to the control (saline solution) during the daily treatment, starting from the first day of damage [10].

The effect of the "Santil" ointment on the resolution of inflammation in long-term unhealed wounding was investigated. The research was carried out on macrophages, isolated from wounds treated with ointment or petrolatum (a comparison drug), and implanted in mice. A significant increase in pro-reparative and a decrease in pro-inflammatory macrophages polarization in both acute inflammatory process and chronic diabetic wound, have been revealed. Wound macrophages in the ointment-treated group showed an increased production of anti-inflammatory cytokines IL-10 and TGF- β , as well as a reduced production of pro-inflammatory cytokines TNF- α and IL-1 β . Wound treatment with clostridial collagenase attenuated the transactivation of factor NF-KB and significantly reduced STAT6-phosphorylation. These results, make it possible to consider collagenase as a potential anti-inflammatory agent that can be effective in chronic wound inflammation, including diabetic wounds [31]. There are numerous data on the effectiveness of the "Santil" ointment, used as a wound healing agent in clinical practice. The data confirm that the ointment based on collagenase, is safe and effective for the treatment of skin ulcers and burn wounds, helping to reduce the healing time and the severity of pain symptoms, reduce the risk of infection. The effectiveness of the "Santil" ointment for surgical treatment of diabetic foot ulcers, bedsores and trophic varicose ulcers, as well as burns, has been demonstrated in a number of clinical trials in various institutions (for example, inpatient, outpatient and long-term kinds of care) [32–34].

The data of clinical studies of the "Moricrol" ointment containing *Paralithodes camtschaticus* collagenase, were registered. The use of the ointment in patients with hard-to-heal skin wounds and the wounds after skin transplantation, contributed to the engraftment of transplants in patients and the absence of rough deform-

ing scars. A reduction of the healing time of extensive wounds in the mucosa of the oral cavity when using the "Moricrol" ointment, was revealed. It has been noted that the treatment with "Moricrol" in purulent wounds, contributed to a faster wound surface cleansing [35].

In clinical research it was found out, that the combined application of wound coating "Multiferm" (the collagenase *Paralithodes camtschaticus* and chitosan complex), photodynamic and NO-therapy, accelerated the defect reparation in the case of trophic ulcers. It was manifested in accelerate purifying of purulent necrotic content от гнойно-некротического содержимого and the granulation formation 2.4 times faster. The average healing time also decreased by an average of 9.2 days [36].

Clinical efficacy of various collagenase-based anti-scar agents has been studied. The effectiveness of collagenases in the scar therapy is associated with their ability to hydrolyze collagen excess [37–41]. For example, the possibility of hypertrophic scars correction by means of electrophoresis in a solution of weak electrolytes with the "Polycollagenase-K" preparation has been investigated. During the treatment, the analysis of the scars state was carried out by EHF-dielectrometry method. The tendency of the moisture content increase in the scar tissue has been revealed. It was associated with an increase in the fraction of intracellular structured water. It has been detected that the hydration of the scar tissue under the influence of "Polycollagenase-K", approached the values peculiar to those of healthy skin at the similar localization. According to the authors' data, the increase in the moisture content in the scar tissues, was due to the water release during the collagen hydrolysis. Alongside with the destruction of the excess collagen, the normalization of microcirculation was noted [8, 39].

The anti-scar activity of the collagenolytic complex from sea stars has been proved: the ability to influence the reduction of collagen gel, the activity of matrix metalloproteinases (MMP), the release of hydroxyproline and the regulation of the activity of fibroblast genes. It was found out, that the complex significantly inhibited the reduction of collagen gel after 2 days of incubation. The expressed activity of MMP-2 and MMP-9 was revealed, which was manifested in the form of a large amount of hydroxyproline release. The fibroblast cell culture treatment significantly reduced fibrocyte proliferation in 3-day cultures. The ability to influence the expression of genes controlling the inflammatory response in fibroblasts, has been established [42].

The clinical studies of the anti-scar properties of the "Moricrol" ointment, detected a decrease in the manifestations of redness, itching and a sense of tightening of keloid scars. In 46 patients, a significant improvement in the condition of the skin, namely paling of tissues and some flattening of the scar tissue in the scar area was revealed. Some softening of the scar and a decrease in the turgor of the scar skin on palpation. was assessed How-

ever, in some patients (18 patients), no tissue softening occurred, and in 5 patients with chronic burn scars, the expected effect was absent [35].

There are research data on gel (group 1 – applications and group 2 – phonophoresis) and a solution (group 3 – electrophoresis) for scars correction with “Fermencol” containing *Paralithodes camtschaticus* collagenase in patients with hypertrophic and keloid scars of different origins. A statistically significant slowdown in the scar growth, paresthesia and itching reducing, scar thickness reducing, the disappearance of inflammation signs was detected when using the gel and solution “Fermencol” after the treatment course of 10-15 days. It was revealed that the use of therapeutic electrophoresis and ultrasound with “Fermencol”®, made it possible to significantly increase the drug intake into the skin [41].

The influence of electro- and ultraphonophoresis “Fermencol” on clinical manifestations of pathological scars in 89 patients with hypertrophic and keloid skin scars, has been investigated. The scars treatment with “Fermencol” electro- and ultraphonophoresis, contributed to a more significant dynamics of clinical signs, compared with “Contractubex” and “Lidase” electro – and ultraphonophoreses. The maximum reduction in clinical manifestations – type, consistency, color and sensitivity of the scar – was observed under the “Fermencol” ultraphonophoresis. The defibrosing effect was most evident with the combination of the drug and the ultrasound exposure [43].

A new method of treatment and prevention of hypertrophic and keloid scars, which implies lidase injection into the thickness of the scar tissue, followed by wet coating containing chitosan and collagenase application to the scars surface in the course of 10-15 procedures, has been developed, patented and clinically tested.

This method’s clinical approbation showed a significant improvement in both – the clinical picture and the patient’s life quality – in relation to the comparison course (collagenase was administered transdermally in therapeutic doses, and lidase was administered intra-dermally by the course of 10-15 daily procedures) [44].

The method of skin scars after the acne correction by means of a course treatment with the “Fermencol” electrophoresis. In group 1, applications of the «Fermencol» gel were used twice a day, in the 2nd group, electrophoresis of the fermencol solution was carried out. A positive result in 86.4% of patients of group 1 and 94.1% of patients of group 2 was revealed: slowing down of the active growth, regression of the scar, disappearance of unpleasant subjective sensations, alignment of the color of the scar and surrounding tissues [45].

In clinical studies, the evaluation of the treatment of hypertrophic scars on patients after surgical interventions on the thyroid gland. The treatment was carried out by collagenase in the form of the dry “Collalysin” powder (collagenase *Clostridium histolyticum*), mixed up with petrolatum. The positive effect, was observed in a month from

the beginning of the treatment by this drug. Collagenase treatment has shown the results similar to the administration of triamcinolone, a hormonal anti-inflammatory drug, into the scar [46]. An emulsion-based composition of “Collalysin” has also been developed for the prevention and treatment of hypertrophic and keloid scars resulting from burns, dermabrasion and plastic surgery [47].

Modern collagenase-based products: assortment in the Russian pharmaceutical market, opportunities for improvement

In the literature data, there is information about the developed and patented medicines based on collagenase. So, the “Collagenase KK” preparation was produced in the form of the lyophilized powder for the application to damaged tissues or aqueous solutions prepared ex tempore and used for wetting wipes or tampons [6, 17, 48]. The disadvantage of this form is the following: the activity of proteolytic enzymes, when directly introduced into the wound, lasts 15-30 minutes, making the therapy ineffective because of inactivating the enzymes. The process is as also characterized by a the relative complexity [49].

Some hydrophilic gels have been developed, e.g., a polyethylene oxide gel with collase [50, 51]. To include *Paralithodes camtschaticus* collagenase, a composition consisting of a mixture of vinylglutarate, vinyl acetate and vinyl alcohol, which turns into a gel-like state when interacting with the wound contents, was used by I. Yu. Sakharov et al. [24]. The ointments on lipophilic bases were offered. So, the ointment with collagenase from *Clostridium histolyticum* was based on petrolatum. The moricrase “Moricrol” ointment on the lipophilic basis of “Eikonal” (a mixture of fatty acids and vitamins A, E, D and F) is known [19]. Currently, these drugs are not available on the pharmaceutical market [6, 52].

The analysis of the literature data showed that currently, collagenase-based products are available for the preparation of solutions for injection and electrophoresis forms, ointments, creams and medical dressings, in the form of powders [4–8, 52–55].

The following commercial drugs are produced abroad: the “Iruksol” and “Santil” ointments produced by “Smith&Nephew”. They include collagenase C. *histolyticum* as a proteolytic complex. It should be noted that these drugs are not registered in the Russian pharmaceutical market [10, 52].

General Residual Life of the System (GRLS) – Encyclopedia of drugs – presents information about the “Collalysin” drug (the INN “Collagenase”), LSR-005615/09 is a proteolytic agent, an enzyme preparation obtained from the culture of *C. histolyticum*. “Collalysin” has a keloidolytics effect. It is available in the form of a powder-lyophilizate dosage form and strength from 100 KE to 1000 KE for a solution for injections and electrophoresis preparation and is recommended for treatment of burns, correction of scars, etc. [6, 52].

“Fermencol” (Russia) is a cosmetic product based on the enzymes of collagenase hydrobionts, designed for the correction of scars. It is available in the form of gel and a set for preparing an electrophoresis solution [54, 55].

A self-absorbable biological dressing “Digestol”, (Russia) is a wound set (RCF 2008/02946). It contains the sum of collagenolytic trypsin-like proteases (collagenases) of *Paralithodes camtschaticus*. It is recommended for the use in purulent and infected wounds, bedsores, trophic ulcers, burns, a diabetic foot syndrome. Dissolving in the wound contents, the wound coating releases the enzyme in the active form. A necrolytic effect of the wound dressing is combined with an anti-inflammatory and regeneration activating effect due to the presence of collagen [8].

Collagenase is also included in the cosmetic product – a dry “Karipain Plus” balm, containing a complex of enzymes: papain, bromelain and collagenase, intended for the treatment of scars [56].

Thus, the Russian pharmaceutical market currently has a small number of collagenase-based products: a lyophilized powder for the preparation of an injection solution (“Collalysin”), recommended for the scars treatment; a medical dressing with collagenase, recommended for the treatment of wounds and necrotic tissues “Digestol”); cosmetic gel and powder “Fermencol”, a dry balm “Karipain Plus”, recommended for the scars treatment. The drugs are represented by only powder-lyophilizate for the preparation of injection solutions and electrophoresis.

A pharmaceutical development of external drugs with proteolytic enzymes, in addition to justifying the optimal dosage form, makes provision for an ointment base or carrier, that ensure the enzyme stability. The analysis of the scientific and technical literature database as well as patents showed the following methods used: enzyme stabilization, the use of lipophilic base, immobilization on a polymer carrier [16, 24, 57, 58].

So, to preserve the activity of the enzyme in the drug, it is possible to use stabilizers, for example, salts. In particular, ammonium sulfate in a certain concentration reversibly inactivates proteolytic enzymes by precipitation, preventing their autolysis. Sulfate ions interact with positively charged amino acids, giving the protein molecule a more compact shape, making it less soluble. This method of enzyme stabilization is implemented in the “Fermencol” gel [59].

In the ointments with collagenase from *C. histolyticum*, petrolatum is used (“Irujol”, “Santil”). Based on petroleum jelly and paraffin, the veterinary ointment “Iruovetin” containing collagenase, was previously produced. The use of these bases is due to the fact that lipophilic components do not contain water, which is a medium for the enzymes autolysis and reproduction of microorganisms-destroyers, which makes it possible to preserve the activity of enzymes for a long time [3, 4, 25].

But as a base, petroleum jelly is characterized by a number of drawbacks: an occlusive effect and a low osmotic activity, which, for a favorable repair, can negatively affect the wound state requiring oxygen and an exudate outflow..

The ointments based on petrolatum, cause inconvenience, i.e., they are poorly washed off with water, because of the pronounced viscosity they are distributed over the surface of the skin with effort. In this regard, the search for the bases devoid of these shortcomings, but at the same time ensuring the preservation of the enzyme activity during the storage period, can be considered a hot topic [5, 52].

Oleogels, e.g. based on Aerosil, can be considered promising bases for preparations of proteolytic enzymes. Thanks to the gel-like structure, gels are easily applied and distributed over the skin. An additional advantage of this base. can be considered the fact, that Aerosil exhibits a high sorption capacity against the decay products of tissues, toxins, microorganisms. The oleogel containing silicones, has also been proven to be effective as a scars treatment [60, 61].

Currently, in external medicines, immobilized enzymes are used on polymer carriers. Immobilization makes it possible to limit the activity of proteolytic enzymes by the damaged area and increases their stability in the wound environment. Thus, when using the enzymes immobilized on fiber-forming carriers, the terms of purification and wound healing are significantly reduced, the consumption of drugs is also reduced in comparison with the free enzymes use [62, 63].

Preparation of fibrous materials with proteolytic enzymes by covalent immobilization, providing activation of the fiber surface, in particular by treatment with oxidants to obtain dialdehyde cellulose, can serve an example, Active aldehyde groups interacting with functional groups of the enzyme to form covalent bonds, are formed on the fiber surface resulting in its fixation [63, 64].

For example, in the “Multiferm” dressing, a copolymer on the basis of dialdehydecellulose treated with chitosan with immobilized enzyme complex of *Paralithodes camtschaticus* hepatopancreas was used. However, one of the problems of chitosan-containing textile dressings, is the “keratinization” of the dressings edges, associated with structural changes in chitosan during immobilization, sterilization and storage. Currently, studies to obtain more stable preparations of immobilized enzymes with improved functional characteristics, are being conducted in this direction [7, 65].

One of the options for improving the properties of textile dressings, is conferring atraumatic characteristics provided by water-repellent impregnation, e.g., ointments or gel coatings. E.g., a hydrophobic ointment base (the “Branolid” dressing) or wax (“Voscopran”) prevent sticking to the wound and traumatization of the granulations in the process of dressing changes. Changing such a dressing is painless for the patient [66].

Based on this, the research on the creation of atraumatic wipes with immobilized collagenase on a hydrophobic basis can be considered perspective, in our opinion.

CONCLUSION

Collagenases are among the most effective proteolytic enzymes, because of their ability to provide the cleavage of collagen, the main component of wounds and scars. In addition, there are data from preclinical and clinical studies confirming, that collagenases of various origins, alongside with their necrolytic activity, are able to accelerate the process of reparation. Biochemical distinctions of collagenase influence on the wound healing process, as well as biochemical aspects of collagenase anti-inflammatory effect, have been investigated. The results of clinical studies of anti-scar properties of collagenases of different origins confirming their effectiveness in this pathology, have been registered. One of the available raw resources of collagenolytic enzymes is currently the hepatopancreas of crustaceans, in particu-

lar, *Paralithodes camtschaticus*. Due to the composition peculiarities, the enzymes derived from *Paralithodes camtschaticus* hepatopancreas, have a wide specificity as they hydrolyze both native collagen and other protein substrates (gelatin, casein, fibrinogen and serum albumin).

On the pharmaceutical market of the Russian Federation, the range of products with collagenase is not wide, so its expansion can be relevant. The collagenase derived from *Paralithodes camtschaticus* hepatopancreas, can be used as the most affordable raw material with pronounced collagenolytic properties. As the dosage form, more convenient in application and production, gels have their set of advantages. Thanks to their potential sorption properties, the oleogels based on aerosil, can be considered a promising base for external remedies with collagenolytic enzymes. The development of gels and atraumatic dressings with collagenase obtained from *Paralithodes camtschaticus* as the most affordable raw materials, can be considered significant for practical pharmacy.

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All authors had equally contributed to the research work.

CONFLICTS OF INTEREST

The authors and peer reviewers of this paper report no conflicts of interest.

REFERENCES

- Mayorova AV, Sysuev BB, Hanalieva IA, Vihrova IV. Modern assortment, properties and perspectives of medical dressings improvement of wound treatment. *Pharmacy & Pharmacology*. 2018;6(1):4–32. doi:10.19163/2307-9266-2018-6-1-4-32.
- Paramonov BA. Kollagenoliticheskie fermenty Chast' 2. Primenenie dlya ochishcheniya ran [Collagenolytic enzymes Part 2. Application for wound cleansing] *Kosmetika i meditsina*. 2016;(2):38–48. Russian.
- Demidova-Rice TN, Hamblin MR, Herman IM. Acute and impaired wound healing: pathophysiology and current methods for drug delivery. Part 1. Normal and chronic wounds: biology, causes, and approaches to care. *Adv Skin Wound Care*. 2012;25(7):304–314. DOI:10.1097/01.ASW.0000418541.31366.a3
- McCallon, SK, Weir D, Lantis JC. Optimizing Wound Bed Preparation With Collagenase Enzymatic Debridement. *J Am Coll Clin Wound Spec*. 2015;15;6(1–2):14–23. doi: 10.1016/j.jccw.2015.08.003.
- Veterinarnye preparaty v Rossii. Spravochnik. [Veterinary drugs in Russia. Handbook] Moscow: Rus Videl'; 2017: 448 p. Russian.
- Entsiklopediya Lekarstv [Encyclopedia Of Drugs]. RLS. Issue 25 / edited by G. L. Vyshkovsky. Moscow: VEDANTA; 2016: 1288 p. Russian.
- Wound coatings and consumables. Multiform. [Internet] Available from: <http://poliform.ru/multiform>. [cited 2019 May 15] (date accessed: 15.05.2019)
- Digesta [Internet]. Available from: http://www.mazi.ru/izdeliya_naruzhnogo_primeneniya/digestol.html. [cited 2019 May 15]
- Paramonov BA. Kollagenoliticheskie fermenty. Chast' 1. Nereshennye i spornye voprosy teorii i praktiki [Collagenolytic enzymes. Part 1. Unsolved and controversial issues of theory and practice]. *Kosmetika i meditsina*. 2016; 1: 32–41. Russian.
- Sheets AR, Demidova-Rice TN, Shi L, Ronfard V, Grover KV, Herman IM. Identification and Characterization of Novel Matrix-Derived Bioactive Peptides: A Role for Collagenase from Santyl® Ointment in Post-Debridement Wound Healing? *PLoS One*. 2016; 26, №11(7). doi: 10.1371/journal.pone.0159598.
- Konon AD, Petrovskiy SV, Shamburova MYu, Uvarova AV, Kozlova YuO, Grigoryeva MV, Moskvichev BV. Osobennosti biotekhnologij klostridial'nyh kollagenaz – perspektivnyh fermentov medicinskogo naznacheniya [Features of clostridial collagenase biotechnology – emerging enzymes for medical application]. *Medicine of extreme situation*. 2016;2 (56):45–58. Russian.
- Mozhina NV, Rudenskaya GN. Kollagenoliticheskie fermenty patogennykh mikroorganizmov [Collagenolytic enzymes of pathogenic microorganisms] *Biomeditsinskaya Khimiya*. 2004;50(6):539–553. Russian.
- Artjukov AA, Menzorova NI, Kozlovskaja EP, Kofanova NN, Kozlovskij AS, Rasskazov VA. Fermentnyj preparat iz gepatopankreasa promyslovyykh vidov krabov i sposob ego poluchenija [Enzyme preparation from hepatopancreas

- of commercial crab species and method for production of the same enzyme preparation from hepatopancreas of commercial crab species and method for production of the same]. Russian Federation patent (RF) 2280076, 20.07.2006. Russian.
14. Semenova SA, Rudenskaya GN, Lyutova LV, Nikitina OA. Vydelenie i svoystva izoformy serinovej kollagenoliticheskoj proteinyazy kamchatskogo kraba *Paralithodes camtschatica* [Isolation and properties of serine collagenolytic proteinase isoform of Kamchatka crab *Paralithodes camtschatica*]. Biochemistry. 2008; 73(10): 1403–1413. Russian.
 15. Demina NS, Rototaev DA. Fermentnyj ranozazhivljajushhij preparat [Enzymatic wound healing medication]. Russian Federation patent (RF) 2484811, 20.06.13. Russian.
 16. Kulmetieva MA, Korotaeva AI, Belov AA. Immobilizacija proteoliticheskogo kompleksa iz gepatopankreasa kraba na hitozansoderzhashhie celluloznye nositeli v prisutstvii glicerina [Immobilization of proteolytic complex from crab hepatopancreas on chitosan-containing cellulose carriers in the presence of glycerol]. Advances in Chemistry and Chemical Technology. 2014; XXVIII (5): 30–32. Russian.
 17. Kozlovskaya EP, Artyukov AA, Kozlovskii AS, Vozzhova, Kofanova NN, Elyakov GB. Ranozazhivlyayushchee sredstvo «Kollagenaza KK» shirokogo spektra deistviya. Russian Federation patent (RF) 2093166. 20.10.97. No. 29. Russian.
 18. Belov AA, Filatov VN, Belova EN, Filatov NV. Medicinskaja povjazka, sodержashhaja kompleks proteoliticheskikh fermentov, vključaja kollagenoliticheskie proteazy iz gepatopankreasa kraba [Medical bandage containing proteolytic enzyme complex including collagenolytic proteases from crab hepatopancreas]. Russian Federation patent (RF) 2268751. 27.01.06. No. 3. Russian.
 19. Isaev VA, Lyutova LV, Karabasova MA, Rudenskaya GN, Kupenko OG, Stepanov VM. Sostav dlya lecheniya gnoino-troficheskikh yazv i prolezhnei. Russian Federation patent (RF) 2074709. 10.03.1997. Russian.
 20. Salamone M, Cuttitta A, Seidita G, Mazzola S, Bertuzzie F, Ricordi C, Ghersi G. Characterization of collagenolytic/proteolytic marine enzymes. Chemical engineering transactions. 2012; 27 (1): 1–6.
 21. Daboor SM, Budge SM, Ghaly AE, Brooks S-L, Deepika D. Extraction and Purification of Collagenase Enzymes: A Critical Review. Am. J. Biochem. & Biotech. 2010; (6)4: 239–263.
 22. Sivakumar P, Sampath P, Chandrakasan G. Collagenolytic metalloprotease (gelatinase) from the hepatopancreas of the marine crab, *Scylla serrata*. Comparative Biochemistry and Physiology. Part B. 1999; 123: 273–279.
 23. Zinatullin RM, Khatmullina KR, Gizatullin TR, Kataev VA. Puti povysheniya jeffektivnosti jepitelizacii troficheskikh i dlitel'no ne zzhivajushhij ran [Ways to improve epithelialization of trophic and non-healing wounds]. Bashkortostan Medical Journal. 2013; 8(6): 109–111. Russian.
 24. Vernikovskiy VV, Stepanova EF. Immobilizovannye proteazy dlya ochishcheniya ranevykh poverkhnostei. Russian Journal of General Chemistry. 2010; LIV (6): 94–100.
 25. Voronkov AV, Stepanova EF, Zhidkova YY, Gamzeleva OY. Sovremennye podhody farmakologicheskoy korrrekcii patologicheskikh rubcov. [Modern approaches of pharmacological correction of pathological scars]. Fundamental research. 2014;3–2:301–308. Russian.
 26. Sakharov IYu, Litvin FE, Mit'kevich OV. Gidroliz belkov kollagenoliticheskimi proteinazami kamchatskogo kraba. Bioorganicheskaya khimiya. 1994;20(2):190–195.
 27. Payne WG, Salas RE, Ko F. Enzymatic debriding agents are safe in wounds with high bacterial bioburdens and stimulate healing. Eplasty. 2008; 8: R. e17
 28. Riley Kathleen N, Herman Ira M. Collagenase promotes the cellular responses to injury and wound healing in vivo. Journal of burns and wounds. 2005;4: 112–124.
 29. Isaev VL, Lyutova LV, Karabasova MA, Kupenko OG, Andreenko GV, Rudenskaya GN. Ranozazhivlyayushchee deistvie mazi s morikrazoi. Voprosy meditsinskoi khimii. 1994; 40(3):46–48.
 30. Ivankova YuO, Stapanova EF. Razrabotka mazi reparativnogo dejstviya s kollagenazoj kamchatskogo kraba. Advances in current natural sciences. 2014;8: 161–162.
 31. Amitava Das, Soma Datta, Eric Roche, Scott Chaffee, Elizabeth Jose1, Lei Shi, Komel Grover, Savita Khanna, Chandan K. Sen, Sashwati Roy Novel mechanisms of Collagenase Santyl Ointment (CSO) in wound macrophage polarization and the resolution of wound inflammation. [Internet]. 2018;8: 1696. DOI:10.1038/s41598-018-19879-w [cited 2019 March 1] Available from: <https://www.ncbi.nlm.nih.gov/pubmed/>
 32. Waycaster C, Carter MJ, Gilligan AM, Mearns ES, Fife CE, Milne CT. Comparative cost and clinical effectiveness of clostridial collagenase ointment for chronic dermal ulcers. [Internet]. J Comp Eff Res. 2018; 7(2): 149–165. doi: 10.2217/ce-2017-0066. [cited 2019 March 1] Available from: www.ncbi.nlm.nih.gov/pubmed/29076747
 33. Shi L, Carson D. Collagenase Santyl ointment: a selective agent for wound debridement. [Internet]. J Wound Ostomy Continence Nurs. 2009;36:S12–6. doi: 10.1097/WON.0b013e3181bfdd1a. [cited 2019 March 1] Available from: <https://www.ncbi.nlm.nih.gov/pubmed/19918145>
 34. Pham CH, Collier ZJ, Fang M, Howell A, Gillenwater TJ. The role of collagenase ointment in acute burns: a systematic review and meta-analysis. [Internet]. Wound Care. 2019;28(1) (Sup 2): S9–S15. doi: 10.12968/jowc.2019.28.Sup2.S9. [cited 2019 March 1] Available from: <https://www.ncbi.nlm.nih.gov/pubmed/30767636>
 35. Rudenskaya GN, Lyutova LV, Karabasova MA, Andreenko GV, Isaev VA, Brusov AV, Badnina EI, Reznikova AE, Ageeva LV. Lechebnoe dejstvie mazi morikrol. Moscow University Chemistry Bulletin. 2000; 41(6): 414–416.
 36. Lucevich OE, Tamrazova OB, Kuleshov IYu, Sorokatiy AA, Shikunova Alu, Usmonov UD, Starichkov IG. Vozdushno-plazmennye potoki v rezhime koagulyacii, no-terapii v kompleksnom lechenii dlitel'no nezazhivajushhij i hronicheskij ran (jazv) nizhnij konechnostej [Air-plasma flow in the mode of coagulation, NO-therapy in complex treatment of prolonged unhealed and chronic wounds (ulcer) of the lower extremities] Moscow Surgical Journal. 2011;2 (18): 9–13.
 37. Karpova TN, Matytsin VO. Ocenka jeffektivnosti primeneniya sredstva «Fermenkol» v celjah profilaktiki i korrrekcii rubcov. Fizioterapevt. 2008; 6: 53–54. Russian.
 38. Olejnik GA, Grigor'eva TG, Korkunda SV, Cogoev AA Opyt

- ispol'zovaniya preparata «Fermenkol» v profilaktike i lechenii patologicheskikh rubcov // Vestnik neotlozhnoj i vosstanovitel'noj mediciny. 2014; 15(1): 90. Russian.
39. Paramonov BA, Turkovskij II, Antonov SF, Klimova OV, Semenov DP, Bondarev SV. Razrushenie izbytochnogo vnekletochного матрикса как составляющaja lechenija patologicheskikh rubcov kozhi (ocenka v opytah in vitro) [Surplus intracellular matrix destruction as part of pathological skin scars healing (in vitro experiments estimation)] Vestnik Jesteticheskoy Mediciny 2009; 8(3): 69–73. Russian.
 40. Paramonov BA., Turkovskij II., Antonov SF, Klimova OV, Semenov DP, Bondarev SV. Fermentnaja terapija patologicheskikh rubcov kozhi [Pathological skin scars enzyme therapy]. Vestnik neotlozhnoj i vosstanovitel'noj mediciny. 2009; 8(2). 24–28. Russian.
 41. Chasnoits ACh, Zhilinski EV, Serabrou AE, Tsimashok NYu. Ocenka protivorubcovoj jeffektivnosti preparata Fermenkol [Antiscar efficiency evaluation of Fermenkol®]. Mezhdunarodnye obzory: klinicheskaja praktika i zdorov'e. 2016; 1(19): 24–34. Russian.
 42. Zhi Jiang Li, Sang Moo Kim The Application of the Starfish Hatching Enzyme for the Improvement of Scar and Keloid Based on the Fibroblast-Populated Collagen Lattice. Applied Biochemistry and Biotechnology. 2014; 173(4): 989–1002.
 43. Karpova TN, Ponomarenko GN, Samtsov AV. Elektro- i ul'trafonoforezkollagenazy v korrekcii rubcov kozhi [Electro- and ultraphonophoresis of collagenase for correction of dermal scars] Vestnik Rossijskoj voenno-medicinskoj akademii. 2009; (1): 89–94. Russian.
 44. Zinatullin RM, Gil'manov AZh, Khunafin SN, Simonova ES. Sposob lecheniya i profilaktiki razvitiya keloidnykh i gipertroficheskikh rubtsov. Russian Federation patent (RF) 2220741, 10.01.2004. Russian.
 45. Shimanskaya IG, Volotovskaya AV. Metody korrekcii rubcovykh izmenenij kozhi u pacientov na fone ugrevoj bolezni [Methods of correction of cicatricial skin changes in patients in case of acne] Medicinskie novosti. 2015; 9: 38–40. Russian.
 46. Trunin EM, Kandalova IG, Nyn' IV, Berestovaja LK, Obrezkova AV. Ispol'zovanie Kollalizina dlja lechenija gipertroficheskikh rubcov posle operacij na shhitovidnoj zheleze. Poliklinika. 2009; (1): 120–121. Russian.
 47. Zamylova TI, Karakosova TA, Stepanova ZV. Sredstvo for the prevention and treatment of hypertrophic and keloid scars. Russian Federation patent (RF) 2114603, 10.07.1998. Russian.
 48. Stonik V.A. Morskie prirodnye coedinenija. put' k novym lekarstvennym preparatam. Actanaturae. 2009; 2: 16–27. Russian.
 49. Percev IM, Dacenko BM, Gun'ko VG. Mnogokomponentnye mazi na gidrofil'noj osnove. Farmacija. 1990; 39(5): 73–77. Russian.
 50. Ostrovidova GU, Makeev AV. Napravlennoe regulirovanie biologicheskoy aktivnosti mnogokomponentnykh polimernykh struktur. Rossijskij zhurn. priklad. himii. 2002;75(9): 1477–1480. Russian.
 51. Omigov VV, Markovich NA, Balahnin SM, Malygin JeG, Zinov'ev VV, Sandahchiev LS. Morfologicheskaja ocenka vozdejstvija kollagenazy kamchatskogo kraba Parolithodes camtschatica na termicheskij ozhog v jeksperimente. Bjul. jeksper. biol. i mediciny. 1996; 122(7): 97–100.
 52. Gosudarstvennyj reestr lekarstvennyh sredstv [State register of medicines] [Internet]. Moscow, 2019. [cited 2019 April 15] Available from: <http://grls.rosminzdrav.ru>.
 53. Gosudarstvennyj reestr medicinskih izdelij i organizacij (individual'nyh predprinimatelej), osushhestvljajushhih proizvodstvo i izgotovlenie medicinskih izdelij [State register of medical devices and organizations (individual entrepreneurs) engaged in the production and manufacture of medical devices] [Internet]. Moscow, 2019. [cited 2019 April 15] Available from: <http://www.roszdravnadzor.ru/services/misearch>.
 54. Fermenkol [Internet]. [cited 2019 April 15] Available from: <http://fermenkol.ru/fermenkol>.
 55. Peresadina SK, Vasin AS. Primenenie fonoforeza gelja fermentol v lechenii rubcov postakne. Dermatology in Russia. 2017; S. 1: 75–76. Russian.
 56. Fistal' NN. Ocenka jeffektivnosti preparata «Karipain Pljus» v lechenii posleozhogovykh rubcov. Poliklinika. 2012; 4–1: 118–119. Russian.
 57. Raspopova E.A., Korotaeva A.I., Malenko O.E., Belov A.A. Kinetika termoinaktivacii proteoliticheskogo kompleksa iz gepatopankreasa kraba, stabilizirovannogo polisaharidnymi soedinenijami [Kinetics of thermal inactivation of proteolytic complex from crab hepatopancreas, stable polysaccharide compound]. Fundamental research. 2013; 11 (part 4): 656–661. Russian.
 58. Perlamutrov JuN, Olkhovskaya KB. The effectiveness of the cream containing stabilized hyaluronidase for the correction of cicatricial skin changes [Jeffektivnost' krema, soderzhashhego stabilizirovannuju gialuronidazu, dlja korrekcii rubcovykh izmenenij kozhi] Dermatologija. Prilozhenie k zhurnalu CONSILIUM MEDICUM. 2017; 1: 5–9. Russian.
 59. Klimova OA. Method of transdermal introduction of polypeptides into the body. Russian Federation patent (RF). 2462265 (13), 27.09.2012. No. 27.
 60. Ivankova J.O., Vernikovskij V.V., Stepanova E.F. Issledovanie po vyboru osnovy dlja naruzhnoj lekarstvennoj formy kollagenazy. [Study selection framework for external medicinal forms collagenase] Modern problems of science and education. 2015;(2):478.
 61. Astrahanova M.M. Izuchenie reologicheskikh svojstv i vysvobozhdenija iz ajerosilsoedzhashhih mazednykh osnov. Farmacija. 1981; 29(6): 28–31.
 62. Sysuev BB, Akhmedov NM, Samoshina EA, Zaleskih DS, Ivanilova MA, Samarskaya AA, Barbarosh OS. Sovremennye aspekty primeneniya nanotekhnologii pri razrabotke lekarstvennyh form novogo pokolenija [Modern aspects of the application of nanotechnology in the development of a new generation of dosage forms (review)]. Drug development and registration. 2015;3(12):88–96.
 63. Belov AA, Belova EN, Filatov VN. Tekstil'nye materialy, soderzhashhie hitozan i proteoliticheskij kompleks iz gepatopankreasa kraba, dlja medicinskih celej [The textile materials containing chitosan and proteolytic complex from hepatopancreas of the crab, for the medical purposes]. Biomeditsinskaya Khimiya. 2009; 55(1):61–67.
 64. Raspopova EA, Korotaeva AI, Malenko OE, Belov AA. Kine-

- tika termoinaktivacii proteoliticheskogo kompleksa iz gepatopankreasa kraba, stabilizirovannogo polisaharidnymi soedinenijami [Kinetics of thermal inactivation of proteolytic complex from crab hepatopancreas, stable polysaccharide compound]. Fundamental research. 2013;11 (P. 4):656–661.
65. Shurshina AS, Kulish EI, Kolesov SV, Zakharov VP. Preparation of Enzyme-Containing Chitosan Films Pharmaceutical Chemistry Journal. 2015; 49(3): 196–198.
66. Veselov AJe. Opyt ispol'zovaniya ranevyh pokrytij «Voskopran®», «Parapran®», «Voskoposorb®», «Gelepran®» v kompleksnom lechenii detej s ozhogovoj travmoj. Medicinskaja sestra. 2008;(3): 33.

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